

INHIBITION OF TGF β SIGNALING AND ITS IMPLICATIONS IN ANTICANCER TREATMENTS

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The transforming growth factor- β (TGF β) is a potent regulator of tumorigenesis. In cancer, two distinctive behaviors of TGF β have been reported as a tumor suppressor at early stage of the disease, and as a tumor promoter at later stages. The past decades, the dualistic role of TGF β has garnered a lot of attention. As a result, cancer researchers' has been tasked to elucidate how TGF β signaling may lead to metastatic dissemination, how to tackle carcinogenesis and which therapeutic strategies should be adopted. Consequently, TGF β signaling pathways have been considered as appropriate targets for cancer therapy. The TGF β therapeutic strategies have emerged at three levels: ligand, ligand-receptor interaction and intracellular signaling level. Promising inhibitors of TGF β signaling have entered clinical trials and shown encouraging results. Here we review the three strategies of TGF β signaling inhibition and their applications in treatment of cancer.

Key Words: TGF β , cancer, inhibition of TGF β signaling, therapeutic strategy.

The TGF β is a potent cytokine endowed with remarkable functionalities allowing it to perform multiple tasks. Among these tasks, there are regulation of cell proliferation, differentiation, and apoptosis. The cell type and the cell environment may influence the function of the cytokine, enabling it to control multitudinous processes either normal or pathological. As examples of physiological events related to TGF β signaling are embryogenesis, wound healing and tissue homeostasis. Regarding pathological disorders, such as cancer, arteriosclerosis, fibrosis and Marfan's syndrome, compiling evidences have shown that loss of control of the TGF β signaling is associated with these conditions. In the neoplastic transformation, TGF β plays two conflicting roles of a tumor suppressor and a tumor promoter. The inhibition of TGF β signaling pathways may be achieved at three levels in the TGF β signaling pathways. The first strategy is to target the TGF β ligand. The second strategy is to affect the interaction between the TGF β ligand and the TGF β receptors. Finally, the third strategy focuses on the receptor-mediated signaling cascade. There is a considerable diversity of inhibitors designed to approach each of the three levels. These inhibitors have already shown different beneficial aspects in preclinical and clinical studies.

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Abbreviations used: Abs – antibodies; ALK5 see T β R-I; ASO – antisense oligonucleotides; EMT – epithelial-to-mesenchymal transition; mAbs – monoclonal antibodies; miRNA – micro interfering RNA; RNAi – RNA interference; R-Smads – receptor-regulated Smads; SARA – Smad anchor for receptor activation anchorage; siRNA – short interfering RNA; Smad – small mothers against decapentaplegic; TGF β – transforming growth factor beta; T β R-I – type I transforming growth factor receptor; T β R-II – type II transforming growth factor receptor; T β R-III – type III transforming growth factor receptor; T β Rs – TGF β receptors; T-cells – T lymphocytes; TF – transcription factors.

TGF β : STRANGE CASE OF “TGF β ” DR JEKYLL OR MR HYDE?

Carried out by Robert W. Holley in the early 70s, the initial study leading to the discovery of TGF β and its naming as a *transforming* growth factor were based on its ability to induce malignant behavior of normal fibroblasts. This brought the idea that TGF β might be a key factor in transformation of cells [1, 2]. Meanwhile, other experiments indicated a conflicting function of TGF β on cells, that of a tumor suppressor [3]. Today, it is well established that conceded roles to the cytokine are cancer stage dependent [3, 4]. In the early stage of cancer development, TGF β can suppress tumor growth, whereas in the late-stage it can take on role of a tumor promoter, favoring spreading of metastasis [5]. TGF β is assumed to arbitrate a broad range of physiological processes e.g. wound healing, proliferation, epithelial homeostasis, embryogenesis and apoptosis but also pathological processes such as Marfan's syndrome, fibrosis, carcinogenesis including angiogenesis and epithelial-to-mesenchymal transition (EMT) [6–8]. Even three decades after its discovery, it is a difficult task to ascribe one single role to the TGF β in the case of carcinogenesis. The “whimsical” behavior of this cytokine leads to the conclusion that it might be both, a kind of “Guardian angel” by its ability to inhibit tumor proliferation, but nevertheless a kind of “Devil” by its aptitude to enhance metastasis spreading, and for that reason, would deserve the dual title of “Dr Jekyll/Mr Hyde” [9, 10].

TGF β SIGNALING: “TO SMAD, OR NOT TO SMAD?” [11]

The following model for TGF β signaling pathway through Smad proteins has been suggested in several reports (Fig. 1) [12–17].

The signal is triggered through binding of the mature TGF β ligand to the extracellular domain of type II TGF β receptor (T β R-II) or to the accessory receptor, type III TGF β receptor (T β R-III) which transfers TGF β to T β R-II. Following the transfer of the cytokine to T β R-

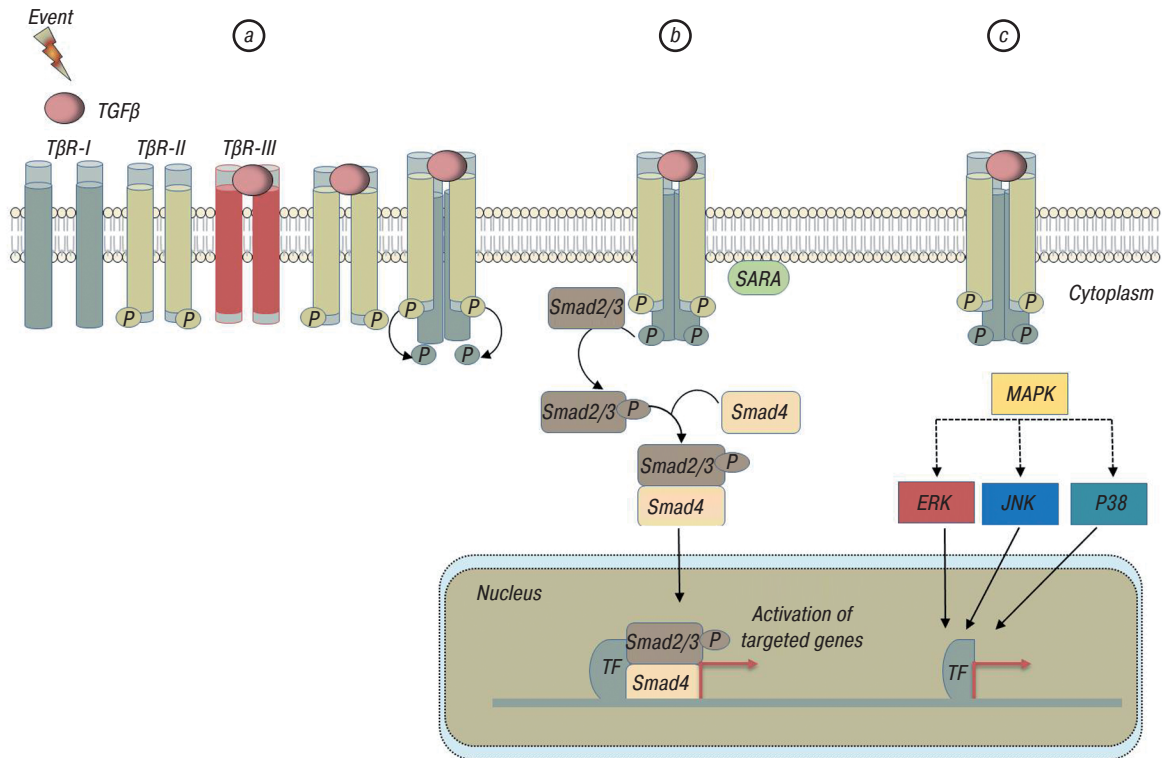


Fig. 1. TGFβ Smad and non-Smad signaling pathways from initiation to nucleus. (a) Signal initiation. After the secretion of TGFβ, the active cytokine can bind TβR-II or the accessory receptor TβR-III which presents it to TβR-II. TGFβ binding to TβR-II leads to bridging of TβR-I into the complex, and allows TβR-I to phosphorylate TβR-II. (b) Smad-dependent pathway. The R-Smads (Smad2 and Smad3) are activated by TβR-I. However their recruitment can be eased by auxiliary proteins, e.g. SARA. These activated R-Smads complex a Co-Smad (Smad4). Finally, the complex is imported into the nucleus, where with the help of other co-operators it regulates expression of targeted genes. (c) Smad-independent pathway. TGFβ can regulate expression of a wide range of genes by inducing other signaling cascade independently of the Smad-dependent pathway, such as shown MAPK

II, the constitutively active receptor TβR-II recruits and phosphorylates the signaling type I TGFβ receptor (TβR-I) [18]. The TβR-I acts downstream of the type II, and determines the specificity of intracellular signals by phosphorylating a subset of transcriptional cytoplasmic factors (TF), major linchpin of the signaling pathway Smad2 and Smad3. Also called R-Smads (receptor-regulated or activated Smads), for the reason that Smad2/3 protein activation is under the control of the receptor TβR-I [13]. Several phosphoisoforms of the R-Smads have been identified as a result of TGFβ and Ras/MAPK pathways activation [19, 20]. The phosphorylation of these first intracellular mediators stimulates their interactions with Smad4 (Co-Smad), a co-mediator and other protein partners e.g. SARA (Smad anchor for receptor activation anchorage), Dab2, Endofin, Axin, etc. [10, 21]. Once assembled, the complex Smad4-Smad2/3 shuttles to the nucleus where it may exert distinct transcriptional control [22, 23]. Besides this Smad-dependent pathway which is doubtless the most-well known TGFβ signaling pathway, other pathways have been identified, clustered and termed as Smad-independent pathways [24, 25]. These other signaling cascades can be activated by TGFβ, and in such a way can orchestrate the transcription of target genes. Among them are described various branches of the MAPK, Rho-like GTPase and PI3K/AKT pathways [24–28]. It is broadly accepted that during carcinogenesis, the Smad-dependent pathway correlates with the

anti-proliferative or tumor suppressor functions of TGFβ, and that the Smad-independent pathways are involved in TGFβ pro-malignant functions [29].

THREE APPROACHES TO INHIBIT TGFβ SIGNALING

Taking account of TGFβ involvement in carcinogenesis (tumor suppression and tumor promotion), the targeting of TGFβ signaling pathway for therapeutics purposes was an ineluctable choice. By dint of intensive works, over fifteen years, the therapeutic strategies to disrupt TGFβ signaling have emerged at three levels: ligand, receptor-ligand interaction and intracellular signal transduction (Fig. 2). Several inhibitors have entered clinical trials, from phase I to III. The Table 1 summarized the current knowledge on therapeutic strategies to impede TGFβ signaling.

Intervention on the ligand level

The signaling pathway's first component is the ligand. Therefore an interest in targeting the transcriptional products of TGFβ-coding genes in order to restrain the synthesis of TGFβ has been applied. The technique called Gene Silencing by RNA Interference (RNAi) allows regulation of the gene expression. The srNAi technology is based on two types of small molecules of RNA: the micro interfering RNA (miRNA) and the short interfering RNA (siRNA) [30]. These small molecules act by binding complementary sequences on specific mRNAs, therefore preventing translation and in that way silencing TGFβ genes. Binding of an-

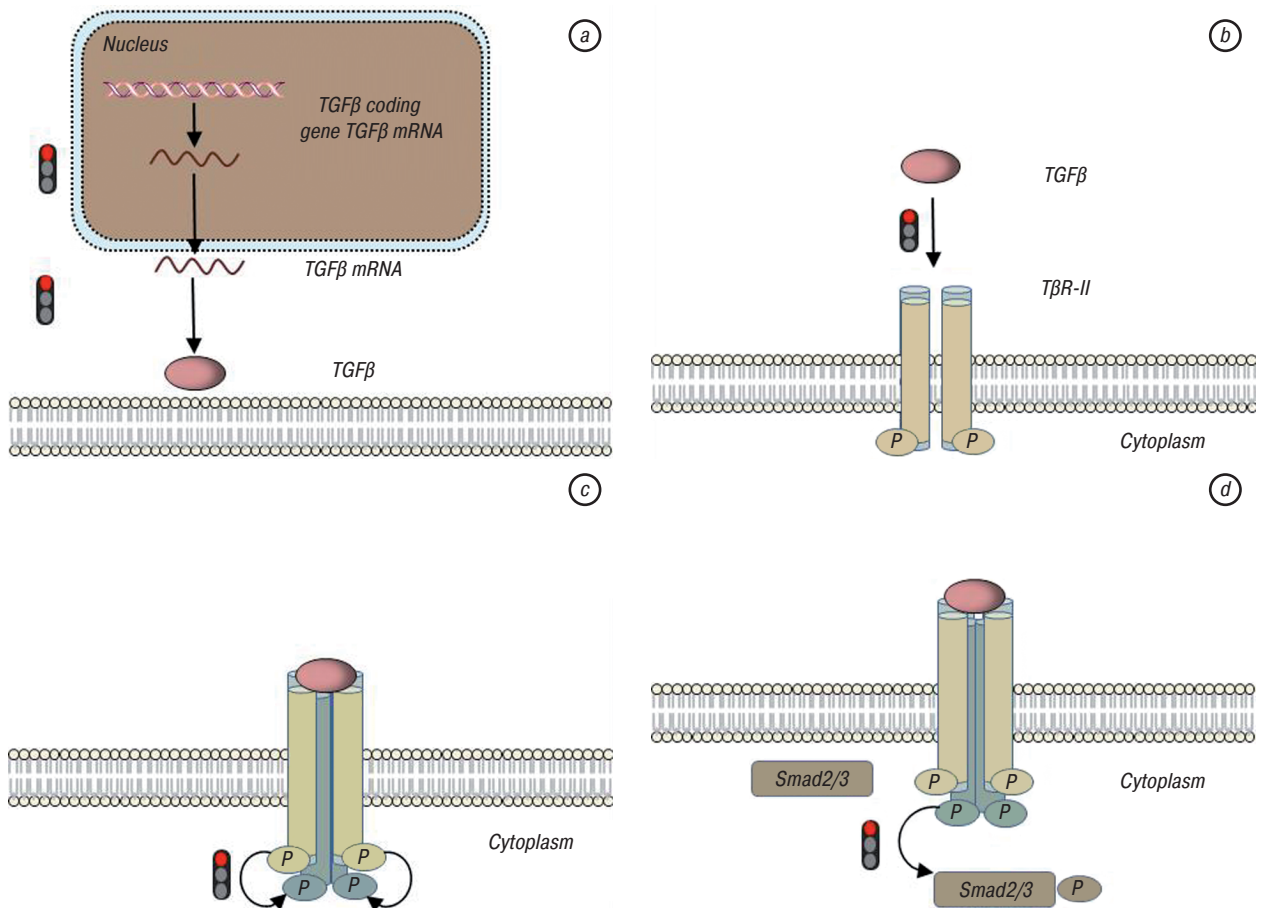


Fig. 2. Current strategies to impede TGFβ signaling. (a) Direct or indirect inhibition of TGFβ secretion. The first measure consists to prevent the secretion of TGFβ directly by action on transcript products using mAbs or ASOs or via indirect routes by decreasing the secretion rate of the cytokine. (b) Inhibition of TGFβ-receptor binding. Another therapeutic line lies in the neutralization of the secreted cytokine by mAbs, soluble receptors or natural TGFβ inhibitors, and therefore blocking the ligand-receptor binding. (c) Inhibition of TGFβ receptor activation. Antagonism of the transduction signal through hampering of TβR-I may be achieved by inhibitors interfering with the ATP-binding pocket or the Smad-binding pocket of the kinase. (d) Inhibition of Smad activation. Finally to stymie the progression of TGFβ signaling on the transduction level one may target Smad2/3 directly. Strategies (a), (b) and (c) are currently being explored, and discussed in the text

tisense oligonucleotides (ASO) to RNAs and targeting TGFβ mRNA made silencing of TGFβ gene possible [31, 32]. The trabedersen, also termed AP-12009, from Antisense Pharma is an ASO responsible for silencing of TGFβ2 gene [29]. In recurrent and refractory high-grade glioma patients, promising results have already been obtained and led to the clinical trials phase III [33, 34]. However, in spite of a high specificity, trabedersen administration remains an issue. In high grade glioma patients a neurosurgical intervention is required to set-up a relatively complicated drug delivery system. This drug delivery system includes a pump placed outside the body which is connected to an internal catheter flowing to the brain [35–37]. Currently, Antisense Pharma is performing phases I/II clinical trials in pancreatic neoplasms, melanoma and colorectal neoplasms using intravenous delivery that already show encouraging efficacy [38]. By neutralizing the TGFβ2 mRNA produced by tumor cells, the Belagenpumatumucel-L or Lucanix™ from NovaRx was expected to restore tumor antigen recognition by immune effector cells i.e. T-cells [39]. In patients with non-small cell lung cancer, results from a phase II study suggest that the number of circulating tumor

cells at baseline appears to correlate with the overall survival. Such results highlight that further explorations remain needed [40, 41]. Although, the published reports are not enough clear regarding the adverse effects, this lack of information may have a considerable impact on the future safety status of neutralizing RNAs as drugs [36].

Intervention on the ligand-receptor interaction level

Drugs of the second level target interaction between the ligand and the specific receptor. To date intervention on the ligand-receptor level encompasses three categories of compounds: monoclonal antibodies (mAbs), natural TGFβ inhibitors and soluble TGFβ receptors (fusion constructs). Here we focus on the mAbs due to the broad use in clinic of antibodies as drugs targeting different signaling receptors. The application of mAbs as a therapeutic end (i.e. immunotherapy) for cancer can be explained by their high specificity [42–44]. Several investigators have demonstrated in cancer mice models that a neutralization of the three isoforms of TGFβ circulating in the bloodstream using several mAbs e.g. 1D11 and 2G7 affected the tumor growth [6]. The mAbs list

Table. Overview of therapeutic TGF β signaling inhibitors used in pre-clinical and clinical studies. (A) Direct or indirect inhibition of TGF β secretion. (B) Inhibition of TGF β /receptor binding. (C) Inhibition of TGF β receptor activation. (D) Inhibition of Smad activation

	Target	Generic name	Status	Application	References
(A)	TGF β mRNA	AP-15012	discovery	oncology	29
	TGF β 1 mRNA	AP-11014	adv. preclinical	oncology	58
	TGF β 2 mRNA	AP-12009 (Trabedersen)	III recruiting	oncology	33, 34, NCT00761280
	TGF β secretion	Tranilast	preclinical	various	59, 60–62
	TGF β 2 secretion	Bevacizumab [®] (Avastin)	II recruiting	various	63, NCT00121134 NCT00733408
	TGF β 2	Glionix [™]	II–III initiated	oncology	64, 65
	TGF β 2 secretion	Lucanix [®] (Belagenpumatucl-L)	III recruiting	oncology	40, 41, NCT00676507
(B)	TGF β 1	Metelimumab [®] (CAT-192)	II discontinued	scleroderma	47, 49
	TGF β 2/3	Lerdelimumab [®] (CAT-152/Trabio)	III discontinued	various	46, 49, 66
	Pan TGF β	Fresolimumab [®] (GC-1008)	I	various	45/NCT01284322 NCT01401062
	Pan TGF β	SR-2F	preclinical	oncology	67
	Pan TGF β	1D11	preclinical	oncology	36
	Pan TGF β	2G7	preclinical	oncology	68–70
(C)	T β R-I	A-83-01	preclinical	oncology	71
	T β R-I	GW6604	preclinical	fibrosis	72, 73
	T β R-I	IN-1130	preclinical	fibrosis	74
	T β R-I	Ki26894	preclinical	oncology	75
	T β R-I	LY2157299	I/II	oncology	76, NCT01220271 NCT01246986 NCT01373164
	T β R-I	LY364947 (HTS-466284)	preclinical	various	36, 77
	T β R-I	LY550410	preclinical	various	48
	T β R-I	L Y5 73636-sodium (Tasisulam)	I/II/III suspended	oncology	78, 79
	T β R-I	LY580276	preclinical	various	48
	T β R-I	NPC-30345	preclinical	various	80, 81
	T β R-I	SB-431542	preclinical	oncology	51, 52
	T β R-I	SB-505124	preclinical	various	53
	T β R-I	SD-093	preclinical	oncology	81, 82
	T β R-I	SD-208	preclinical	various	82, 83
	T β R-I	Sml6	preclinical	oncology	84
	T β R-I	SM305	preclinical	fibrosis	85
	T β R-I	SX-007	preclinical	oncology	86
	T β R-I	Antp-Sm2A	preclinical	oncology	57
	T β R-I /II	LY2109761	preclinical	oncology	87
		P144 (Disitertide)	preclinical/II	various	88, 89
	P17	preclinical	various	88, 90	
	T β R-III	sRIII	preclinical	oncology	91, 92
(D)	Smads	Trx-CBP	preclinical	oncology	93
			discontinued		
	Smads	Trx-FoxHlb	preclinical	oncology	93
	Smads	Trx-Lefl	preclinical	oncology	93
	Smad2/3	Trx-SARA	preclinical	oncology	94
Smad3	SIS3	preclinical	fibrosis	95	

Abbreviations: adv. – advanced, NCT – Clinical Trial Registry Numbers. Source: ClinicalTrials.gov

targeting TGF β includes GC1008, CAT-152 and CAT-192 [45–47]. All three Abs are up to date the most developed antibodies in clinical trials [29, 36, 48]. The pilot study and the phase II studies carried out on patients with advanced malignant melanoma or renal cell carcinoma have shown a reasonable tolerance vis-à-vis GC1008 and a neutralization of TGF β , holding promise of a novel cancer therapeutic agent [45]. However several adverse effects have been noticed, such as fatigue, headache, epistaxis, gingival bleeding, skin rash [6, 45]. Nonetheless, a phase II protocol expansion study is recruiting patients with metastatic malignant melanoma. This protocol allows monitoring of the GC1008 effects in blood samples from patients. A phase II study of patients with breast cancer is planned with GC1008 [6]. The CAT-192 or Metelimumab[®] and the CAT-152 or Lerdelimumab[®] are human IgG4 mAbs directed against TGF β 1 and TGF β 2, respectively. Early clinical studies have suggested that Metelimumab[®] was safe and well tolerated, with a long half-life and has completed phase I/II studies with patients with cutaneous systemic sclerosis. No trials in cancer have yet been initiated [49]. It is important

to bear in mind that the antibodies must surpass substantial obstacles to reach the tumor mass [36, 50]. Among them, the physical barrier including vascular endothelium, stromal barriers, high interstitial pressure and epithelial barriers may explain in part why the therapeutic antibodies have a moderated success in treatment of cancer as compared to fibrotic disorders [36, 44, 50].

Intervention on the intracellular signaling level

The transforming factor receptors (T β Rs) are the gateways of the intracellular signaling. Therefore, drugs blocking TGF β receptors intracellular activity have been developed, thus constituting the third group of inhibitors. Most of them are small molecule inhibitors targeting the kinase of T β Rs. However, others inhibitors target Smads interaction with T β R, using peptide aptamers to Smad (Table 1). This section of our report focuses on the inhibitors targeting the T β Rs. Among these inhibitors there are two categories; the small and the large molecules. The currently developed inhibitors may have an imidazole scaffold, such as SB-431542 and SB-505124, or a pyrazole scaffold such as LY-580276 [36, 48]. Most of these inhibitors are

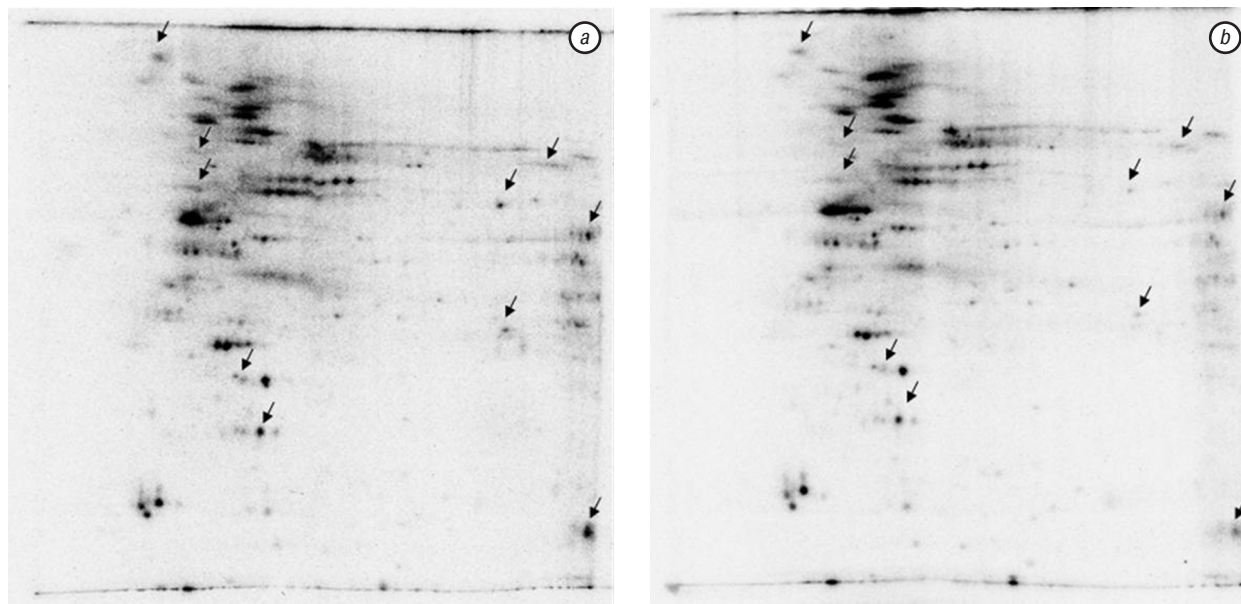


Fig. 3. Screen for specificity of a kinase inhibitor. *a* — without inhibitor; *b* — +SB-431542. ATP-binding site interfering inhibitor SB-431542 was added to cell extract (right 2D gel) or not (left 2D gel), *in vitro* kinase reaction was performed under conditions promoting autophosphorylation of kinases. Proteins were separated, and ^{32}P incorporation was detected after exposure in a PhosphorImagerTM. Arrows show migration positions of proteins which were affected by addition of SB-431542

directed towards T β R-I kinase catalytic ATP-binding site [48]. Two representative candidates are being developed by GlaxoSmithKline, e.g. SB-431542 and SB-505124. Both aim to fully abrogate or strongly down-regulate the T β R-I mediated signaling cascade. These imidazole-based compounds have already shown remarkable effects at nanomolar concentrations, comprising inhibition of the TGF β -induced Smad phosphorylation, as well as inhibition of a reporter gene [51–53]. Finally, the T β R-I blockade has shown effects on cellular responses such as the cell cycle arrest and EMT of mammary epithelial cells *in vitro* [6, 51]. In contrast to its analog (i.e. SB-431542), SB-505124 has been revealed to be three to five times more potent [53]. However it has been shown that these inhibitors may not be specific to T β R-I. Fig. 3 shows a proteomics screen to evaluate the specificity of SB-431542. This screen detected a number of phosphorylated proteins which were affected by the drug. The assay was designed to detect predominantly auto-phosphorylated kinases. Detection of multiple targets of the drug underscores importance of the unbiased evaluation specificity of the drug. The lack of high specificity could be explained by the inherent analogous structure shared by several kinases on their ATP-binding pocket, e.g. p38, bringing Lahn *et al* to voice the concern that such off-target inhibition might be liable to unexpected toxicity [36]. Finally, even if such molecules present the particularly advantages of an oral administration and a high selectivity, they remain nonetheless not enough specific and should be carefully monitored in future clinical trials [54–56]. This is in view of certain cases of resistance to the drug, or side-effects such as cardiac conditions reported in the literature [55, 56].

Whilst some researchers have focused in recent years to create inhibitors targeting the ATP-binding site, another approach to target the kinase on the

substrate-binding site has been reported [57]. This novel strategy aims to inhibit signaling by blocking the substrate-binding site of the T β R-I kinase with peptides mimicking the Smad2. This new class of inhibitors acts as “decoys” which once occupying the Smad2-binding pocket, prevent Smad2 phosphorylation, and hence its activation. This idea should by definition allow a high specificity that some ATP-mimicking inhibitors do not offer. So far, only one group has produced and investigated the effects of such compounds. The results have shown that this kind of compounds can indeed disturb TGF β signaling by blockade of T β R-I *in vivo* and *in vitro*, in Mv1Lu cells. On top of this, there have been shown that those new inhibitors affected TGF β 1-dependent phosphorylation of endogenous Smad2, as well as gene stimulation. Finally, these pseudo-substrates have shown higher efficiency vis-à-vis the T β R-I kinase than to kinases of other type I receptor of TGF β and BMP family [57]. Nowadays, investigations on normal and cancer celllines are ongoing. In view of the encouraging results, it is clear that development of pseudo-substrate inhibitors may lead to new therapeutic strategies to impede TGF β signaling.

CONCLUSION

As metastasis dissemination remains the major cause leading to death of cancer patients, significant efforts have been undertaken over the years to tackle cancer by blockade or at least by decreasing development of the metastasis. To face this challenge, the inhibition of TGF β signaling appears as a therapeutic strategy. This strategy has been approached at three levels: ligand, receptor-ligand binding and intracellular signaling levels. Among them, the ASOs and the mAbs technologies are the most advanced. Yet, the inhibitors likely to experience a fast growth will undoubtedly be the small-molecules drugs. Novel type of inhibi-

tors, e.g. substrate-mimicking drugs, requires further developmental efforts. However, already now we have examples of successful application of inhibition of TGF β signaling for the benefit of cancer patients. This ensures that TGF β inhibitors came into anticancer treatment to stay. The inhibitors have been tested in a number of assays, and have shown their efficiency. However, developmental work requires much more. Taking into account potential benefit for patients and results of clinical trials with other types of TGF β signaling inhibitors, there is a strong support to continue development of the substrate-mimicking inhibitors.

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REFERENCES

- Holley RW. A unifying hypothesis concerning the nature of malignant growth. *Proc Natl Acad Sci USA* 1972; **69**: 2840–1.
- Moses HL, Roberts AB. The discovery of TGF- β : a historical perspective. In: *The TGF- β Family*. Derynck R, Miyazono K, eds. New York: Cold Spring Harbor Laboratory Press, 2008: 1–28.
- Roberts AB, Anzano MA, Wakefield LM, *et al.* Type beta transforming growth factor: a bifunctional regulator of cellular growth. *Proc Natl Acad Sci USA* 1985; **82**: 119–23.
- Witz IP. Yin-Yang activities and vicious cycles in the tumor microenvironment. *Cancer Res* 2008; **68**: 9–13.
- Massagué J. TGF β in cancer. *Cell* 2008; **134**: 215–30.
- Tan AR, Alexe G, Reiss M. Transforming growth factor- β signaling emerging stem cell target in metastatic breast cancer. *Breast Cancer Res Treat* 2009; **115**: 1–63.
- Dumont N, Arteaga CL. Transforming growth factor- β and breast cancer: tumor promoting effects of transforming growth factor- β . *Breast Cancer Res* 2000; **2**: 125–32.
- Ikushima H, Miyazono K. TGF β signalling: a complex web in cancer progression. *Nat Rev Cancer* 2010; **10**: 415–24.
- Roberts AB, Wakefield LM. The two faces of transforming growth factor in carcinogenesis. *Proc Natl Acad Sci USA* 2003; **100**: 8621–3.
- Muraoka-Cook RS, Dumont N, Arteaga CL. Dual role of transforming growth factor B in mammary tumorigenesis and metastatic progression. *Clin Cancer Res* 2005; **11**: 937–43.
- Kowanetz M. Novel regulators of the TGF- β signaling pathway. *Acta Universitatis Upsaliensis*. Digital comprehensive summaries of Uppsala dissertations from the Faculty of Medicine 57. 2005; 69pp. ISBN 91–554–6303–7.
- Shi Y, Massagué J. Mechanism of TGF- β signaling from cell membrane to the nucleus. *Cell* 2003; **113**: 685–700.
- Wrana JL, Attisano L, Wieser R, *et al.* Mechanism of activation of the TGF- β receptor. *Nature* 1994; **370**: 341–7.
- Attisano L, Wrana JL. Signal transduction by the TGF- β superfamily. *Science* 2002; **296**: 1646–7.
- Massagué J. TGF- β signal transduction. *Annu Rev Biochem* 1998; **67**: 753–91.
- Moustakas A, Heldin CH. The regulation of TGF β signal transduction. *Development* 2009; **136**: 3699–714.
- Feng XH, Derynck R. Specificity and versatility in TGF- β signaling through Smads. *Annu Rev Cell Dev Biol* 2005; **21**: 659–93.
- Elliott RL, Blobe GC. Role of transforming growth factor beta in human cancer. *J Clin Oncol* 2005; **23**: 2078–93.
- Matsuzaki K. Smad phosphoisoform signaling specificity: the right place at the right time. *Carcinogenesis* 2011; **32**: 1578–88.
- Ikushima H, Miyazono K. TGF- β signal transduction spreading to a wider field: a broad variety of mechanisms for context-dependent effects of TGF- β . *Cell Tissue Res* 2012; **347**: 37–49.
- Kang JS, Liu C, Derynck R. New regulatory mechanisms of TGF- β receptor function. *Trends Cell Biol* 2009; **19**: 385–94.
- Nakao A, Imamura T, Souchelnytskyi S, *et al.* TGF- β receptor-mediated signalling through Smad2, Smad3 and Smad4. *EMBO J* 1997; **16**: 5353–62.
- Dennler S, Itoh S, Vivien D, *et al.* Direct binding of Smad3 and Smad4 to critical TGF β -inducible elements in the promoter of human plasminogen activator inhibitor-type 1 gene. *EMBO J* 1998; **17**: 3091–100.
- Derynck R, Zhang YE. Smad-dependent and Smad-independent pathways in TGF- β family signalling. *Nature* 2003; **425**: 577–84.
- Zhang YE. Non-Smad pathways in TGF- β signaling. *Cell Res* 2009; **19**: 128–39.
- Parvani JG, Taylor MA, Schiemann WP. Noncanonical TGF- β signaling during mammary tumorigenesis. *J Mammary Gland Biol Neoplasia* 2011; **16**: 127–46.
- Mu Y, Gudey SK, Landström M. Non-Smad signaling pathways. *Cell Tissue Res* 2012; **347**: 11–20.
- Moustakas A, Heldin CH. Non-Smad TGF- β signals. *J Cell Sci* 2005; **118**: 3573–84.
- Nagaraj NS, Datta PK. Targeting the transforming growth factor- β signaling pathway in human cancer. *Expert Opin Investig Drugs* 2010; **19**: 77–91.
- Harel-Bellan A. Silence, clean up genes! *Med Sci (Paris)* 2006; **22**: 993–4.
- Ait-Si-Ali S, Guasconi V, Harel-Bellan A. RNA interference and its possible use in cancer therapy. *Bull Cancer* 2004; **91**: 15–8.
- Chan, JHP, Lim S, Wong WSF. Antisense oligonucleotides: from design to therapeutic application. *Clin Exp Pharmacol Physiol* 2006; **33**: 533–40.
- Bogdahn U, Hau P, Brawanski A, *et al.* Specific therapy for high-grade glioma by convection enhanced delivery of the TGF- β 2 specific antisense oligonucleotide AP12009. *Proc Am Soc Clin Oncol* 2004; **23**: 110.
- Bogdahn U, Hau P, Stockhammer G, *et al.* Targeted therapy for high-grade glioma with the TGF- β 2 inhibitor trabedersen: results of a randomized and controlled phase IIb study. *Neuro Oncol* 2011; **13**: 132–42.
- Antisense Pharma. 1st June 2011; Press release.
- Lahn M, Kloeker S, Berry BS. TGF- β inhibitors for the treatment of cancer. *Expert Opin Invest Drugs* 2005; **14**: 629–41.
- Misra A, Ganesh S, Shahiwala A, *et al.* Drug delivery to the central nervous system: a review. *J Pharm Pharm Sci* 2003; **6**: 252–273.
- Antisense Pharma. 7th June 2011; Press release.
- PR Newswire. 4th November 2009; Press release.
- Nemunaitis J, Dillman RO, Schwarzenberger PO, *et al.* Phase II study of Belagenpumatucel-L, a transforming growth factor beta-2 antisense gene-modified allogeneic tumor cell vaccine in non-small-cell lung cancer. *J Clin Oncol* 2006; **24**: 4721–30.

41. Nemunaitis J, Nemunaitis M, Senzer N, *et al.* Phase II trial of Belagenpumatucel-L, a TGF- β 2 antisense gene modified allogeneic tumor vaccine in advanced non-small cell lung cancer (NSCLC) patients. *Cancer Gene Ther* 2009; **16**: 620–4.
42. Waldmann TA. Immunotherapy: past, present and future. *Nature Med* 2003; **9**: 269–77.
43. Chames P, Baty D. Bispecific antibodies for cancer therapy: the light at the end of the tunnel? *MAbs* 2009; **1**: 539–47.
44. Stewart TJ, Smyth MJ. Improving cancer immunotherapy by targeting tumor-induced immune suppression. *Cancer Metastasis Rev* 2011; **30**: 125–40.
45. Morris JC, Shapiro GI, Tan AR, *et al.* Phase I/II study of GC1008: A human anti-transforming growth factor- β (TGF β) monoclonal antibody (MAb) in patients with advanced malignant melanoma (MM) or renal cell carcinoma (RCC). *J Clin Oncol* 2008; **26**: 9028.
46. Khaw P, Grehn F, Holló GP, *et al.* A phase III study of sub-conjunctival human anti-transforming growth factor β (2) monoclonal antibody (CAT-152) to prevent scarring after first-time trabeculectomy. *Ophthalmology* 2007; **114**: 1822–30.
47. Denton CP, Merkel PA, Furst DE, *et al.* Recombinant human anti-transforming growth factor I antibody therapy in systemic sclerosis: a multicenter, randomized, placebo-controlled phase I/II trial of CAT-192. *Arthritis Rheum* 2007; **56**: 323–33.
48. Yingling JM, Blanchard KL, Sawyer JS. Development of TGF- β signalling inhibitors for cancer therapy. *Nat Rev Drug Discov* 2004; **3**: 1011–22.
49. Foley S. CAT may abandon skin drug after trial results disappoint. 10th February 2004; *The Independent* (London).
50. Christiansen J, Rajasekaran AK. Biological impediments to monoclonal antibody-based cancer immunotherapy. *Mol Cancer Ther* 2004; **3**: 1493–501.
51. Halder SK, Beauchamp RD, Datta PK. A specific inhibitor of TGF- β receptor kinase, SB-431542, as a potent antitumor agent for human cancers. *Neoplasia* 2005; **7**: 509–21.
52. Laping NJ, Grygielko E, Mathur A, *et al.* Inhibition of transforming growth factor (TGF)- β 1-induced extracellular matrix with a novel inhibitor of the TGF- β type I receptor kinase activity: SB-431542. *Mol Pharmacol* 2002; **62**: 58–64.
53. DaCosta Byfield S, Major C, Laping NJ, *et al.* SB-505124 is a selective inhibitor of transforming growth factor- β type I receptors ALK4, ALK5 and ALK7. *Mol Pharmacol* 2004; **65**: 744–52.
54. Fabian MA, Biggs WH 3rd, Treiber DK, *et al.* A small molecule-kinase interaction map for clinical kinases inhibitors. *Nat Biotechnol* 2005; **23**: 329–36.
55. Noble MEM, Endicott JA, Johnson LN. Protein kinase inhibitors: insights into drug design from structure. *Science* 2004; **303**: 1800–5.
56. Orphanos GS, Ioannidis GN, Ardavanis AG. Cardiotoxicity induced by tyrosine kinase inhibitors. *Acta Oncol* 2009; **48**: 964–70.
57. Yakymovych I, Engström U, Grimsby S, *et al.* Inhibition of transforming growth factor β signaling by low molecular weight compounds interfering with ATP- or binding sites of the TGF β type I receptor kinase. *Biochemistry* 2002; **41**: 11000–7.
58. Schlingensiepen KH, Bischof A, Egger T, *et al.* The TGF- β 1 antisense oligonucleotide AP 11014 for the treatment of non-small-cell lung, colorectal and prostate cancer: pre-clinical studies. *Proc Am Soc Clin Oncol* 2004; **23**: 227.
59. Izumi K, Mizokami A, Li YQ, *et al.* Tranilast inhibits hormone refractory prostate cancer cell proliferation and suppresses transforming growth factor β 1-associated osteoblastic changes. *The prostate* 2009; **69**: 1222–34.
60. Chakrabarti R, Subramaniam V, Abdalla S, *et al.* Tranilast inhibits the growth and metastasis of mammary carcinoma. *Anticancer Drugs* 2009; **20**: 334–45.
61. Subramaniam V, Chakrabarti R, Prud'homme GJ, *et al.* Tranilast inhibits cell proliferation and migration and promotes apoptosis in murine breast cancer. *Anticancer Drugs* 2010; **21**: 351–61.
62. Subramaniam V, Ace O, Prud'homme GJ, *et al.* Tranilast treatment decreases cell growth, migration and inhibits colony formation of human breast cancer cells. *Exp Mol Pathol* 2011; **90**: 116–22.
63. Lee SH, Leem HS, Jeong SM, *et al.* Bevacizumab accelerates corneal wound healing by inhibiting TGF- β 2 expression in alkali-burned mouse cornea. *BMB Rep* 2009; **42**: 800–5.
64. Ge L, Hoa N, Bota D A, *et al.* Phase I clinical trial of a TGF- β antisense-modified tumor cell vaccine in patients with advanced glioma. *Cancer Gene Ther* 2006; **13**: 1052–60.
65. Ge L, Hoa N, Bota DA, *et al.* Immunotherapy of brain cancers: the past, the present, and future directions. *Clin Dev Immunol* 2010; **2010**: 296453.
66. Grehn F, Holló GP, Khaw P, *et al.* Factors affecting the outcome of trabeculectomy: an analysis based on combined data from two phase III studies of an antibody to transforming growth factor β 2, CAT-152. *Ophthalmology* 2007; **114**: 1831–8.
67. Yang YA, Dukhanina O, Tang B, *et al.* Lifetime exposure to a soluble TGF- β antagonist protects mice against metastasis without adverse effects. *J Clin Invest* 2002; **109**: 1607–15.
68. Ganapathy V, Ge R, Grazioli AV, *et al.* Targeting the transforming growth factor- β pathway inhibits human basal-like breast cancer metastasis. *Mol Cancer* 2010; **9**: 122.
69. Arteaga CL, Hurd SD, Winnier AR, *et al.* Anti-transforming growth factor (TGF)- β antibodies inhibit breast cancer cell tumorigenicity and increase mouse spleen natural killer cell activity. Implications for a possible role of tumor cell/host TGF- β interactions in human breast cancer progression. *J Clin Invest* 1993; **92**: 2569–76.
70. Biswas S, Guix M, Rinehart C, *et al.* Inhibition of TGF- β with neutralizing antibodies prevents radiation-induced acceleration of metastatic cancer progression. *J Clin Invest* 2007; **117**: 1305–13.
71. Tojo M, Hamashima Y, Hanyu A, *et al.* The ALK-5 inhibitor A-83-01 inhibits Smad signaling and epithelial-to-mesenchymal transition by transforming growth factor- β . *Cancer Sci* 2005; **96**: 791–800.
72. de Gouville AC, Boullay V, Krysa G, *et al.* Inhibition of TGF- β signaling by an ALK5 inhibitor protects rats from dimethylnitrosamine-induced liver fibrosis. *Br J Pharmacol* 2005; **145**: 166–77.
73. Laping NJ, Huet S. TGF- β receptor kinase inhibitors for the treatment of fibrosis. In: Smad signal transduction: Smads in proliferation, differentiation and disease. ten Dijke P and Heldin C-H eds. Springer Verlag, 2006: 443–59.
74. Moon JA, Kim HT, Cho IS, *et al.* IN-1130, a novel transforming growth factor- β type I receptor kinase (ALK5) inhibitor, suppresses renal fibrosis in obstructive nephropathy. *Kidney Int* 2006; **70**: 1234–43.
75. Ehata S, Hanyu A, Fujime M, *et al.* Ki26894, a novel transforming growth factor- β type I receptor kinase inhibitor, inhibits *in vitro* bone metastasis of a human breast cancer cell line. *Cancer Sci* 2007; **98**: 127–33.
76. Bueno L, de Alwis DP, Pitou C. Semi-mechanistic modeling of the tumour growth inhibitory effects of LY2157299,

a new type I receptor TGF- β kinase antagonist, in mice. *Eur J Cancer* 2008; **44**: 142–50.

77. Sawyer JS, Anderson BD, Beight DW, *et al.* Synthesis and activity of new aryl- and heteroaryl-substituted pyrazole inhibitors of the transforming growth factor- β type I receptor kinase domain. *J Med Chem* 2003; **46**: 3953–6.

78. Haritunians T, Gueller S, O'Kelly J, *et al.* Novel acyl sulfonamide LY573636-sodium: effect on hematopoietic malignant cells. *Oncol Rep* 2008; **20**: 1237–42.

79. Roxanne Nelson. Lilly Suspends Study of Tasisulam for Metastatic Melanoma. *Medscape Medical News Oncology*.

80. Singh J, Chuaqui CE, Boriack-Sjodin PA, *et al.* Successful shape-based virtual screening: the discovery of a potent inhibitor of the type I TGF- β receptor kinase (T β RI). *Bioorg Med Chem Lett* 2003; **13**: 4355–9.

81. Ge R, Rajeev V, Subramanian G, *et al.* Selective inhibitors of type I receptor kinase block cellular transforming growth factor- β signaling. *Biochem Pharmacol* 2004; **68**: 41–50.

82. Ge R, Rajeev V, Ray P, *et al.* Inhibition of growth and metastasis of mouse mammary carcinoma by selective inhibitor of transforming growth factor- β type I receptor kinase *in vivo*. *Clin Cancer Res* 2006; **12**: 4315–30.

83. Uhl M, Aulwurm S, Wischhusen J, *et al.* SD-208 a novel transforming growth factor β receptor I kinase inhibitor, inhibits growth and invasiveness and enhances immunogenicity of murine and human glioma cells *in vitro* and *in vivo*. *Cancer Res* 2004; **64**: 7954–61.

84. Suzuki E, Kim S, Cheung HK, *et al.* A novel small-molecule inhibitor of transforming growth factor β type I kinase (SM16) inhibits murine mesothelioma tumor growth *in vivo* and prevents tumor recurrence after surgical resection. *Cancer Res* 2007; **67**: 2351–9.

85. Ishida W, Mori Y, Lakos G, *et al.* Intracellular TGF- β receptor blockade abrogates Smad-dependent fibroblast activation *in vitro* and *in vivo*. *J Invest Dermatol* 2006; **126**: 1733–44.

86. Tran TT, Uhl M, Ma JY, *et al.* Inhibiting TGF- β signaling restores immune surveillance in the SMA-560 glioma model. *Neuro Oncol* 2007; **9**: 259–70. Erratum in *Neuro Oncol* 2007; **9**: 465.

87. Melisi D, Ishiyama S, Sclabas GM, *et al.* LY2109761, a novel transforming growth factor β receptor type I and type II dual inhibitor, as a therapeutic approach to suppressing pancreatic cancer metastasis. *Mol Cancer Ther* 2008; **7**: 829–40.

88. Vicent S, Luis-Ravelo D, Antón I, *et al.* A novel lung cancer signature mediates metastatic bone colonization by a dual mechanism. *Cancer Res* 2008; **68**: 2275–85.

89. Digna Biotech, ISDIN. 25th March 2010; Press release.

90. Gil-Guerrero L, Dotor J, Huijbregtse IL, *et al.* *In vitro* and *in vivo* down-regulation of regulatory T cell activity with a peptide inhibitor of TGF- β 1. *J Immunol* 2008; **81**: 126–35.

91. Bandyopadhyay A, Zhu Y, Cibull ML, *et al.* A soluble transforming growth factor beta type III receptor suppresses tumorigenicity and metastasis of human breast cancer MDA-MB-231 cells. *Cancer Res* 1999; **59**: 5041–6.

92. Bandyopadhyay A, Zhu Y, Malik SN, *et al.* Extracellular domain of TGF β type III receptor inhibits angiogenesis and tumor growth in human cancer cells. *Oncogene* 2002; **21**: 3541–51.

93. Cui Q, Lim SK, Zhao B, *et al.* Selective inhibition of TGF-responsive genes by Smad-interacting peptide aptamers from FoxH1, Lef1 and CBP. *Oncogene* 2005; **24**: 3864–74.

94. Zhao BM, M.F Hoffman MF. Inhibition of transforming growth factor- β 1-induced signaling and epithelial-to-mesenchymal transition by the Smad-binding peptide aptamer Trx-SARA. *Mol Biol Cell* 2006; **17**: 3819–31.

95. Jinnin M, Ihn H, Tamaki K. Characterization of SIS3, a novel specific inhibitor of smad3, and its effect on transforming growth factor- β -1-induced extracellular matrix expression. *Mol Pharmacol* 2006; **69**: 597–607.